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# Phosphate solubilising inoculants on (Jump start) growth parameters of rice (*Oryza sativa* L.)

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#### Abstract

Phosphorus plays a significant role in several physiological and biochemical activities in plants. Phosphorus in soils is immobilized due to formation of insoluble complexes such as iron and aluminium hydrous oxides and calcium carbonate. Phosphate-solubilizing microorganisms (PSM) play an important role in insoluble phosphates into soluble forms involves processes of acidulation, ion chelation and exchange reactions. Present study, a field experiment was carried out to study the Growth parameters of phosphate solubilising inoculants (JumpStart) on rice variety CO 47 in rice crop. The present study revealed that the treatment  $P_3S_3$  (100% P + JumpStart 10E5) was recorded increased Seedling Growth will be increase in  $P_3S_3$  treatment. The Plant Height will be increased at grain filling (111.67 cm), Root/ Shoot ratio (0.494 mg g<sup>-1</sup>) and Root Volume will be increased at Active tillering and 50% flowering stage. The uptake of phosphorus by plants was found higher in  $P_3S_3$  treatment than control. This might be due to the significant increment of major Growth Parameters such as Root and Shoot ratio by the application of 100% recommended phosphorus with seed treatment of JumpStart 10E5.

Keywords: Phosphorus, rice, growth parameters

#### Introduction

Phosphorus is an important nutrient required by rice (Kim et al., 1998)<sup>[13]</sup> and it has a defined role in part of the plants, Meiosis, Phospholipids and reproductive parts, metabolisms such as root development, photosynthesis, nutrient transport within the plant, (Rasipour and Asgharzadeh, 2007)<sup>[20]</sup>. The judicious and proper use of phosphorous in rice markedly increases the yield and quality of rice. Without adequate supply of plant with phosphorous, plant cannot attain its maximum yield. In the world low level of P is one of the major constraints for rice production. This is particularly apparent under upland conditions commonly characterized by poorly fertile, erodible, badly leached, highly acidic, and Pfixing soils, normally with little or no fertilizer applied (IRRI, 1997)<sup>[11]</sup>. P deficiency is identified as under lowland conditions it's a main factor for limiting the performance of modern rice varieties to approach their optimum yields. P deficiency quick remedy for rice field application of P fertilizers. However, inorganic fertilizers are not always available for rice farmers. Besides, some rice soils having low level of P can also fix it into a highly less soluble mineral. Dobermann et al., (1998)<sup>[4]</sup> resulted that more than 90% of the P fertilizer are not easily available to plants. The microorganisms perform an important role in agriculture by supplying nutrients to plants and reduce the demand of chemical fertilizers (Cakmakci et al., 2006)<sup>[2]</sup>.

The use of P solubilizing microorganisms improves the soil fertility and increase the crop production to fulfil the requirement. *Penicillium bilaii* (a phosphorus solubilising fungus) is a newly isolated soil fungus that has profound effect on solubilisation of phosphorus. The efficacy of *Penicillium bilaii* has been test verified in various crops such as wheat, canola, chick pea, mustard and lentil elsewhere. However, the usefulness of *Penicillium bilaii* has not been tested in rice crop under machine transplanting system. The present study was conducted at field level to evaluate the performance of different inoculation of *Penicillium bilaii* hildii in combination with three levels of phosphatic fertilizer on rice crop.

#### **Materials and Methods**

The Present investigation was carried out in the field no. H7a of Department of Rice, TNAU, Coimbatore during Kharif season of 2015. Rice variety CO 47 seed material collected from Department of Rice. Before sowing, three bags (1 kg each) of rice seeds were soaked for 24

hours. Seeds are treated with three different population of Phosphate solubilising inoculants – *Penicillium bilaii* (Jump Start) at the rate of 160 mg per Kg of seeds and 6 ml of water was added and wait for one minute as per the treatments and compared with the recommended seed treatment of Azophos at the rate of 20 g kg<sup>-1</sup> of seeds and control (No seed treatment). The experiment was conducted at field condition.

The field experiment was laid out in split plot design with three replications. The treatments are as follows three main plot of three different P levels P<sub>1</sub>- 0% (0 kg P ha<sup>-1</sup>), P<sub>2</sub>- 50% P (25 kg P ha<sup>-1</sup>) and P<sub>3</sub><sup>-1</sup>00%P (50 kg P ha<sup>-1</sup>) (Recommended dose). Sub plot of five different seed treatments S<sub>1</sub>- No Seed Treatment, S<sub>2</sub>- JumpStart 10E4 (0.00967 mg kg<sup>-1</sup> of seeds), S<sub>3</sub>-JumpStart 10E5 (0.0967 mg kg<sup>-1</sup> of seeds) and S<sub>5</sub>- Azophos at the 20g kg<sup>-1</sup> of seeds. Five plants were selected at different growth stages for recording observation.

# Results and Discussion

# **Seedling Growth Parameters**

Seedling parameters i.e., root length, shoot length, root volume, number of roots, root shoot ratio, dry weight of seedling and vigour index acid were observed with different seed inoculations of Pencillium bilaii in our study (Table.1). All the parameters were increased with higher P levels and Pencillium bilaii (JumpStart 10E5). Pencillium bilaii released maximum amount of Pi and produced highest amount of IAA and GA results in increase in all parameters. Reported increased root length, shoot length and vigor index in maize when treated with bacterial strains. However, this study identifies the effects of P. bilaii on gross root morphology (i.e., root length and specific root length) under field conditions in rice crop. In contrary, specific root length of rice seedling was higher in P<sub>3</sub>S<sub>3</sub>, in line with the findings of Molla et al (2001) that seed treatment with co-inoculation of Azospirillum and Bradyrhizobium increase the SRL of soybean.

The influence of seed inoculants on seedling growth parameters like Shoot length  $S_3$  (18.4 cm), Root length  $S_3$ (14.2 cm), Root Volume  $S_3$  (20.0 cc), No of roots seedling  $S_3$  (18.0), Root/ Shoot ratio  $S_3$  (0.92), Total Dry Weight  $S_3$ (0.37 mg), Specific Root Length (401.4), Vigour Index (7.40), are presented in (Table.1) and found to be influenced by different seed inoculants. Among the different seed inoculants, (*Penicillium bilaii*) recorded higher values followed by  $S_2$ ,  $S_4$ ,  $S_5$  and  $S_1$ .

#### Plant height

Plant height is considered as an important trait related to growth and development of a plant. The crop length was significantly increased at different crop stages when the seed was inoculated with *Pencillium bilaii* (JumpStart), a phosphours solubilizing fungus. 100% P fertilizer in

combination with S<sub>3</sub> (JumpStart 10E5) have recorded increase in plant height of 20.95% at grain filling stage compared to control. The increase in plant height might be due to the increase activity of meristematic cells when phosphorus application is increased. Increase in shoot and root length may also be attributed to the production of plant growth promoting substances like IAA, GA and cytokine like substances by solubilizing microorganism. Increased cell elongation and multiplication due to enhanced nutrient uptake by plants following inoculation of P solubilizing microorganisms might have caused the increase in plant height. Richa et al. (2007) reported positive correlation with phosphorus and plant height. The co-inoculation of Azospirillum and PSB significantly increased the shoot height of foxtail millet over the control. In contrary, Mehrvarz (2008) <sup>[15]</sup> reported that there is no effect of chemical fertilizer and phosphorus soluble microorganism in plant height.

#### **Root parameters**

Root characteristics such as root/shoot ratio and root volume, of rice plant at different growth stage had revealed the effect of different P levels and seed inoculation. Mehrvarz *et al.* (2008) <sup>[15]</sup> reported that phosphorus solubilizing bacteria increase root morphological characters like root length, and root volume in aerobic rice plants. On the present study among the different treatments,  $P_3S_3$ (100% P + JumpStart 10E5) recorded large influence on root characteristics, which may be due to interaction effect of *Penicillium bilaii* with phosphorus levels. In this study related to maximum root volume was reported in P<sub>3</sub>S<sub>3</sub> over control treatment (Fig.1). These results are in line with the findings rice in chickpea. The neutral or negative response on root-hair frequency with increasing P concentration reported in spinach, rape, tomato and Arabidopsis. These result are supported with the findings. Showed that P starvation, rather than its provision, increased root to shoot dry weight ratios in corn (Zea mays L.) found that root to shoot ratio decreased with increasing P fertilization levels in celery. Treatment  $P_1S_1$  (0.706%) recorded higher root shoot ratio at Active tillering stage (Table 3). Higher root-shoot ratio is often reported for P stressed plants as compared with P sufficient plants. In our study similar to Wissuwa et al. (2005) reported that under P deficiency condition root shoot ratio 20- 30% increase in rice crop. The root growth and development would be more susceptible to declines in soil moisture, nutrients and temperature because of the shallow distribution of the root system in rice (Zhang and Cai, 2005) <sup>[26]</sup>. Kaymak et al. (2008) reported that phosphorus solubilizing bacteria (Bacillus megaterium) increased root parameters like rooting performances, like root length, dry matter content of root in mint plant. Under low - Phosphorus condition lateral root width will be increase in maize crop (Yonghong Xie et al., 2003).

Table 1: Effect of P levels and Penicillium bilaii inoculants on seedling growth characters of rice

Treatments	Shoot length (cm)	Root length (cm)	Root volume (cc)	No. of roots	Root shoot ratio	Specific root length (cm)	Dry weight per seedling (g)	Vigour index
$S_1$	15.3	8.8	12	13.0	0.56	1.7	0.24	4.56
$S_2$	17.2	13.3	18	17.0	0.81	2.6	0.33	6.60
$S_3$	18.4	14.2	20	18.0	0.92	2.8	0.37	7.40
$S_4$	17.1	12.8	17	16.0	0.78	2.4	0.29	5.80
<b>S</b> 5	16.4	12.7	15	15.0	0.72	2.3	0.27	5.40

Mean	16.88	12.36	16.4	15.8	0.758	2.36	0.3	5.952
SEd	0.024	0.034	0.050	0.033	0.0038	0.0072	0.0021	0.001
CD (P= 0.05)	0.054	0.079	0.116	0.075	0.0088	0.0167	0.0049	0.002

S1 - No seed treatment, S2 - JumpStart 10E4 (0.097 g/kg of seed), S3 - JumpStart 10E5 (0.966 g/kg of seed)

S<sub>4</sub> - JumpStart 10E6 (9.662 g/kg of seed), S<sub>5</sub> - Azophos (20 g/kg of seed)

Table 2: Effect of P levels and Penicillium bilaii inoculants on plant height (cm plant<sup>-1</sup>) at different growth stages of rice

Treatments	AT (40 DAS)	PI (55 DAS)	F (85 DAS)	<b>GF (105 DAS)</b>	Mean
$P_1S_1$	42.63	62.12	85.67	92.33	70.69
$P_1S_2$	46.30	68.60	90.33	100.00	76.31
$P_1S_3$	48.59	70.72	96.33	104.67	80.08
$P_1S_4$	45.83	65.55	87.33	96.67	73.85
$P_1S_5$	44.55	63.55	86.00	94.67	72.19
$P_2S_1$	45.90	63.00	87.67	96.00	73.14
$P_2S_2$	49.90	68.53	92.00	103.33	78.44
$P_2S_3$	52.53	71.10	99.50	107.33	82.62
$P_2S_4$	47.10	66.27	90.00	97.67	75.26
$P_2S_5$	46.14	64.97	89.00	95.00	73.78
$P_3S_1$	47.06	66.13	92.00	98.33	75.88
$P_3S_2$	51.00	70.93	98.00	106.00	81.48
$P_3S_3$	53.55	74.47	103.33	111.67	85.76
$P_3S_4$	49.35	68.37	97.00	100.33	78.76
P <sub>3</sub> S <sub>5</sub>	48.64	66.55	96.33	99.67	77.80
Mean	47.94	67.39	92.70	100.24	
S at P SEd	0.969	1.415	1.094	1.601	
CD (P=0.05)	1.999	2.920	2.257	3.304	

P<sub>1</sub> - 0% Phosphorus, P<sub>2</sub> -50% Phosphorus, P<sub>3</sub> - 100% Phosphorus

S1-No seed treatment, S2 - JumpStart 10E4 (0.097 g/kg of seed), S3 - JumpStart 10E5 (0.966 g/kg of seed)

S<sub>4</sub> - JumpStart 10E6 (9.662 g/kg of seed), S<sub>5</sub> - Azophos (20 g/kg of seed)

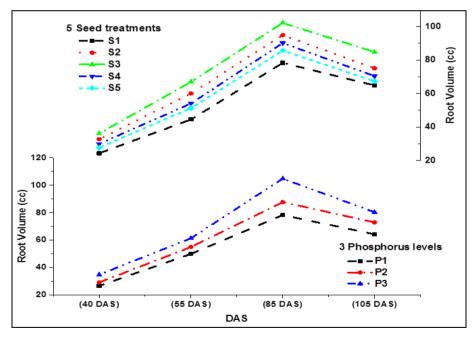


Fig 1: Effect of P levels and Penicillium bilaii inoculants on root volume (cc) at different growth stages of rice

Treatments	AT (40 DAS)	PI (55 DAS)	F (85 DAS)	GF (105 DAS)	Mean
$P_1S_1$	0.706	0.658	0.487	0.434	0.571
$P_1S_2$	0.577	0.583	0.447	0.415	0.506
$P_1S_3$	0.562	0.563	0.438	0.393	0.489
$P_1S_4$	0.591	0.602	0.467	0.400	0.515
$P_1S_5$	0.668	0.591	0.477	0.422	0.540
$P_2S_1$	0.650	0.638	0.476	0.421	0.546
$P_2S_2$	0.576	0.559	0.421	0.400	0.489
$P_2S_3$	0.531	0.533	0.415	0.381	0.465
$P_2S_4$	0.605	0.572	0.438	0.415	0.508
P <sub>2</sub> S <sub>5</sub>	0.638	0.547	0.458	0.428	0.518

$P_3S_1$	0.618	0.524	0.459	0.395	0.499
$P_3S_2$	0.528	0.507	0.411	0.371	0.454
$P_3S_3$	0.494	0.489	0.367	0.359	0.427
P <sub>3</sub> S <sub>4</sub>	0.551	0.546	0.425	0.404	0.482
P <sub>3</sub> S <sub>5</sub>	0.583	0.513	0.436	0.403	0.484
Mean	0.592	0.562	0.441	0.403	
S at P SEd	0.0019	0.0017	0.0012	0.00059	
CD (P= 0.05)	0.0026	0.0023	0.0016	0.00121	

P1 - 0% Phosphorus, P2 - 50% Phosphorus, P3 - 100% Phosphorus

S<sub>1</sub>-No seed treatment, S<sub>2</sub> - JumpStart 10E4 (0.097 g/kg of seed), S<sub>3</sub>-JumpStart 10E5 (0.966 g/kg of seed)

S4-JumpStart 10E6 (9.662 g/kg of seed), S5 - Azophos (20 g/kg of seed)

### Conclusion

I conclude that S<sub>3</sub>- (*Penicillium bilaii* JumpStart 10E5) is the best one compare to other treatment from my research experiment in rice crop.

#### References

- 1. Adesemoye AO, Kloepper JW. Plant-microbes interactions in enhanced fertilizer-use efficiency. Appl Microbiol Biotechnol. 2009;85:1-12.
- Cakmakci R, Donmez F, Aydın A, Şahin F. Growth promotion of plants by plant growth-promoting rhizobacteria under greenhouse and two different field soil conditions. Soil Biol Biochem. 2006;38:1482-1487.
- Degroot CC, Boogaard RVD, Marcelis LFM, Harbinson J, Lambers H. Contrasting effects of N and P deprivation on the regulation of photosynthesis in tomato plants in relation to feedback limitation. J Exp Bot. 2003;54:1957-1967.
- Dobermann A, Cassman KG, Mamaril CP, Sheehy JE. Management of phosphorus, potassium and sulphur in intensive irrigated lowland rice. Field Crop Res. 1998;56:113-138.
- Fales FW. The assimilation and degradation of carbohydrate by yeast cells. J Biol Chem. 1951;193:113-124.
- Gomez KA, Gomez AA. Statistical procedures for Agricultural Research. An IRRI book, Wiley Inter Science publication, John Wile and Sons, New York, USA; c1984. p. 680.
- Hartemink AE, Johnston M, Sullivan JN, Poloma S. Nitrogen use efficiency of taro and sweet potato in the humid lowlands of Papua New Guinea. Agric Ecosyst Environ. 2000;79:271-280.
- 8. Hiscox JD, Israelstam GF. A method for the extraction of chlorophyll from leaf tissue without maceration. Can J Bot. 1979;57:1332-1334.
- 9. Hong Shen, Chen J, Wang Z. Root plasma membrane H+-ATPase is involved in the adaptation of soya bean to phosphorus starvation. J Exp Bot. 2006;57:1353-1362.
- 10. Hooley. Gibberellic acid controls the secretion of acid phosphatase in aleurone layers and isolated aleurone protoplasts of Avena Fatua. J Exp Bot. 1984;35:822-828.
- 11. IRRI (International Rice Research Institute). Annual report for 1972. Los Banos, Philippines; c1997.
- Jeong HL, Chung I, Ryu SS, Park MR, Yun SJ. Differential responses of rice acid phosphatase activities and isoforms to phosphorus deprivation. J Biochem Mol Biol. 2003;36:597-602.
- 13. Kim KY, Jordan D, McDonald GA. Enterobacter agglomerans, phosphate solubilizing bacteria and

microbial activity in soil: effect of carbon sources. Soil Biol Biochem. 1998;89:995-1003.

- Kumar V, Punia SS, Lakshminarayana K, Narula N. Effect of phosphate solubilizing analogue resistant mutants of Azotobacter chroococcum on sorghum. Indian J Agric Sci. 1999;69:198–200.
- Mehrvarz S, Chaichi MR. Effect of Phosphate Solubilizing Microorganisms and Phosphorus Chemical Fertilizer on Forage and Grain Quality of Barley (*Hordeum vulgare* L.). Am-Eur J Agric Environ Sci. 2008;3(6):855-860.
- Muthaiya S. Physiological effects of phosphate inoculants (JumpStart) in maize (*Zea mays* L.). MSc (Agri.) Thesis. Tamil Nadu Agricultural University, Coimbatore; c2010.
- Perderson PL, Carafoli E. Ion motive ATPase. I. Ubiquity, properties and significance to cell function. Trends Biochem Sci. 1987;12:146-150.
- Prasanna R. Physiological impact of phosphate solubilising inoculants (JumpStart) in rice. MSc (Agri) thesis. Tamil Nadu Agricultural University, Coimbatore; c2013.
- Ramezan A, Khavari N, Farzaneh N, Cobra T. Diverse responses of tomato to N and P deficiency. Int J Agric Biol. 2008;1560–8530.
- 20. Rasipour L, Asgharzadeh NA. The interaction between the PSB and Bradirhizobium japanicum growth factors, tumor size and uptake of some nutrients in soybean. Agric Nat Res Sci. 2007;11(5):40-63.
- 21. Ryan MH, Angus JF. Arbuscular mycorrhizae in wheat and field pea crops on a low P soil: increased Zn-uptake but no increase in P-uptake or yield. Plant Soil. 2003;250:225–239.
- 22. Rychter AM, Randall DD. The effect of phosphate deficiency on carbohydrate metabolism in bean roots. Physiol Plantarum. 1994;91:383-388.
- 23. Ullrich CI, Novacky AJ, Fisher E, Liittge U. Relationship between energy dependent phosphate uptake and the electrical membrane-potential in Lemna gibba L. J Plant Physiol. 1984;67:797–801.
- 24. Umbreit WW, Burris RH, Stauffer JF (Eds.). Carbon dioxide and bicarbonate. Manometric techniques. 4th ed. Burgess Publishers; c1964. p. 18-27.
- 25. Wang S, Wu H, Qiao J, Ma L, Liu J, Xia Y, Gao X. Molecular mechanism of plant growth promotion and induced systemic resistance to tobacco mosaic virus by Bacillus sp. Biotechnol. 2009;19(10):1250-1258.
- Xie W, Weng G, Zhang Q. Potential production simulation and optimal nutrient management of two hybrid rice varieties in Jinhua, Zhejiang Province. J Zhejiang Univ Sci. 2007;8(7):486-492.